Review paper

Multidrug resistance

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Since we found verapamil as a multidrug resistance (MDR) reversing agent in 1981, many MDR reversing compounds have been reported. This type of drug must have strong effects with little side effects. We recently found MS-209 and PSC-833 as reversing agents. These two compounds interacted directly with P-glycoprotein, and showed a good MDR reversing effect in vitro and in vivo. MRK16, an antibody against P-glycoprotein, also showed a good therapeutic effect against drug resistant human tumors. MS-209, PSC-833 and the antibody against P-glycoprotein are interesting candidates for clinical use in the future.

Key words: MDR reversing agent, monoclonal antibody, MS-209, PSC-833.

Introduction

Resistance of tumors to a variety of chemotherapeutic agents presents a major problem in cancer treatment. Resistance to such agents as adriamycin (ADM), Vinca alkaloids and actinomycin D can be acquired by tumor cells after treatment with a single drug. The gene responsible for multidrug resistance (MDR), termed *mdr1*, encodes a particular type of membrane glycoprotein (P-glycoprotein) that pumps out various cytotoxic drugs from the cell. The P-glycoprotein has been shown to bind anticancer drugs^{3,4} and to be an ATPase^{5,6} localized at the plasma membrane of resistant cells. Transfection of cloned *mdr1* genes confers MDR on sensitive cells. 8-10

The amount of P-glycoprotein expressed in cells has been measured in tumor samples, and was found to be elevated in intrinsically drug-resistant cancers of the colon, kidney and adrenal, as well as in some tumors that acquired drug resistance after chemotherapy.^{11–15} The expression of the *mdr1* gene in tumors correlates well with clinical drug resistance.^{16,17} In addition to tumor cells, P-glyco-

protein is also expressed in various normal tissues such as adrenal, gravid uterus, kidney, liver, colon and capillary endothelium in brain. 18-24 Pglycoprotein expressed in such normal tissues could have physiologic functions specific to respective tissues.

Because P-glycoprotein appears to be involved in both acquired MDR and intrinsic drug resistance in human cancer, the selective killing of tumor cells expressing P-glycoprotein could be very important for cancer therapy, although the side effects on normal cells expressing P-glycoprotein must be considered carefully. In an effort to devise effective treatments for human drug-resistant cancers, several approaches have been developed to circumvent the MDR of tumor cells, including (i) application of chemosensitizing agents and (ii) application of monoclonal antibodies (mAbs) against P-glycoprotein. These therapeutic approaches are described in this review.

MDR reversing agents

In 1981, we reported that the calcium channel blocker verapamil inhibited active drug efflux and restored drug sensitivity in MDR cells.²⁵ Various compounds, including calcium channel blockers and calmodulin inhibitors, have been shown to enhance the cytotoxic activity of various antitumor agents (refs 25-31, 37; for review, see refs 32 and 33). Verapamil was reported to be a good inhibitor of the photoaffinity labeling of P-glycoprotein with a vinblastine photoanalog^{3,4} and of the transport of Vinca alkaloid into vesicles from MDR cells. 34,35 Direct binding of verapamil to P-glycoprotein was shown using MDR Chinese hamster lung and human K562 cells. 36,37 Verapamil was actively effluxed from resistant cells.³⁷ These findings suggest that verapamil competitively inhibits the transport of antitumor agents by P-glycoprotein, resulting in the reversal of MDR.

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Preliminary clinical studies have been carried out combining calcium channel blockers with chemotherapy. The combination chemotherapy was potentially useful against refractory acute lymphocytic leukemia of children, malignant lymphoma, multiple myeloma, various advanced solid tumors and small cell lung cancer, but induced side effects such as reversible hypotension and arrhythmias.

Cyclosporin A, an immunosuppressive drug, is another chemosensitizer that has been well studied^{38,39} and used for clinical study to overcome drug resistance. Cyclosporin A is also a competitive inhibitor of drug transport by P-glycoprotein. 39 Clinical combination chemotherapy was carried out with cyclosporin A on refractory multiple myeloma patients. 40 Nine out of 21 patients responded to the combination chemotherapy with cyclosporin A. Interestingly, the mdr1 gene was detected in 12 of 15 patients tested before the treatment and seven MDR-positive patients responded to the treatment while no MDR-negative patients did. The clinical studies with verapamil and cyclosporin A suggest that combination chemotherapy with chemosensitizers is potentially effective to overcome clinical drug resistance, especially in patients with resistant tumors expressing P-glycoprotein.

MS-073 is a quinoline derivative that is effective to overcome MDR in vitro and in vivo. 41 MS-073 at 0.1 µM almost completely reversed resistance to vincristine (VCR) in VCR-resistant P388 (P388/VCR) cells in vitro. The compound also reversed the resistance in vitro against VCR, ADM, etoposide and actinomycin D in ADM-resistant human myelogenous K562 (K562/ADM) cells, ADM-resistant human ovarian carcinoma A2780 cells and colchicine-resistant human KB cells. MS-073 administered i.p. daily for 5 days with VCR enhanced the chemotherapeutic effect of VCR in VCR-resistant P388-bearing mice. Increases in life span of 19-52% were obtained by the combination of 100 µg/kg VCR with 3-100 mg/ kg MS-073, as compared with the control. The ability of MS-073 to reverse MDR was higher, especially at low MS-073 doses, than that of verapamil, both in vitro and in vivo. MS-073 enhanced accumulation of [3H]VCR in K562/ADM cells. Photolabeling of Pglycoprotein with 200 nM [³H]azidopine in K562/ ADM plasma membranes was completely inhibited by 10 μM MS-073, indicating that MS-073 reverses MDR by competitively inhibiting drug binding to Pglycoprotein.

FK-506, a novel immunosuppressive agent, was found to be an interesting compound. FK-506 at 3 and $10 \mu g/ml$ completely reversed the resistance

against VCR in vitro in VCR- and ADM-resistant P388 leukemia, and human ovarian cancer A2780 cells. 42 FK-506 also enhanced the cytotoxicity of VCR in highly resistant human K562/ADM in vitro and enhanced the chemotherapeutic effect of VCR in P388/VCR-bearing mice. When 20 mg/kg of the compound was combined with 200 mg/kg of VCR, 151% of the T/C value was obtained. Under the protocol used in this study, FK-506 was more potent than cyclosporin A and verapamil. FK-506 efficiently inhibited [3H]azidopine binding to P-glycoprotein. The binding of VCR to K562/ADM plasma membrane was inhibited by FK-506 as effectively as cyclosporin A. Moreover, the accumulation of VCR in ADM-resistant human ovarian cells (2780AD) was increased by FK-506 as efficiently as cyclosporin A and verapamil.

PSC833 is a non-immunosuppressive analog of cyclosporin A that shows superior effects to cyclosporin A in overcoming MDR *in vitro* and *in vivo*. As a shown to be 10-fold more effective than cyclosporin A *in vitro*. Moreover, PSC833 surpassed cyclosporin A and verapamil in the prolongation of the life span of P388/VCR-bearing mice when chemosensitizers were orally administered with i.p. or i.v. administration of VCR and ADM. PSC833 inhibited [³H]azidopine binding to P-glycoprotein as well as cyclosporin A, and increased cellular accumulation of VCR and ADM in resistant tumors with a lower concentration than cyclosporin A.

MS-209—a quinoline derivative for the reversal of MDR

MS-209, an analog of MS-073, is considered for clinical application because of its efficient oral bioavailability and potentiating effects. 46 We examined the effect of MS-209 on the sensitivity of various MDR cells and their parental cells to VCR. As shown in Table 1, P388/VCR and ADM-resistant P388 (P388/ ADM) cells showed 29- and 40-fold greater resistance, respectively, to VCR as compared with parental P388 cells. When MS-209 was added at a final concentration of 0.1–10 μ M to the MDR P388 cells, MS-209 at 1 and 3 µM completely reversed VCR resistance in P388/VCR and P388/ADM cells, respectively. The sensitivity to VCR in the parental P388 cells was moderately enhanced by MS- 209. MS-209 also remarkably enhanced the sensitivity to VCR in the MDR K562 cells. K562/VCR and K562/ADM cells showed 171- and 1847-fold resistance, respectively, to VCR as compared with parental K562 cells. VCR resistance in MDR K562 cells was completely re-

Table 1. Enhancement of VCR cytotoxicity by MS-209 in MDR cells^a

MS-209 (μM)	IC ₅₀ (ng/ml) of VCR ^b							
	P388	P388/VCR	P388/ADM	K562	K562/VCR	K562/ADM		
0	1.89 (1) ^c	54.5 (1)	75.5 (1)	0.34 (1)	58.2 (1)	628 (1)		
0.1	0.64 (3.0)	19.2 (2.8)	53.9 (1.4)	0.34 (1)	19.5 (3.0)	608 (1.0)		
0.3	0.60 (3.2)	6.26 (8.7)	24.3 (3.1)	0.24 (1.4)	4.04 (14)	433 (1.5)		
1	0.41 (4.6)	1.48 (81)	4.72 (16)	0.20 (1.7)	0.88 (66)	19.3 (33)		
3	0.18 (11)	0.20 (273)	0.92 (82)	0.17 (2.0)	0.55 (106)	2.27 (277)		
10	<0.1 (>19)	` ,	,	, ,	0.21 (277)	0.36 (1744		

^a Tumor cells were seeded in 0.1 ml of culture medium and then treated with graded concentrations of antitumor agents in the absence or presence of MS-209. After 72 h of continuous drug exposure, growth-inhibitory effects were evaluated by MTT assay.

versed by 10 μ M MS-209. A similar potentiating effect of MS-209 on ADM cytotoxicity was observed in the MDR P388 and K562 cells.

We examined the combined chemotherapeutic effect of ADM and MS-209 on P388/ADM-bearing mice. As shown in Table 2, oral administration of MS-209 with ADM given i.p. once a day for five consecutive days starting from day 1 apparently increased the life span of P388/ADM-bearing mice. The most prominent results were obtained when MS-209 was administered with 2 mg/kg ADM. T/C values of 111–144 and 150–194% were obtained with combined treatment

with 50-200 mg/kg MS-209 (experiment 1) and 200-450 mg/kg MS-209 (experiment 2), respectively.

As shown in Figure 1, $[^3H]$ azidopine specifically labeled an M_r 170 000–180 000 protein in K562/ADM cells but not in drug-sensitive K562 cells. In the presence of 100 μ M MS-209, the radiolabeling of P-glycoprotein was completely inhibited. This result suggests that MS-209 directly interacts with P-glycoprotein and inhibits the transport of antitumor agents.

Together with other pharmacological and toxicological profiles (data not shown), the present results

Table 2. Effect of MS-209 on antitumor activity of ADM in P388/ADM-bearing micea

Treatment	n	Median (days)	Range (days)	T/C ^b (%)	Body weight change ^c (g)
		(uays)			
Experiment 1					
control	12	9.0	8–12	100	+1.0
ADM (2 mg/kg)	6	9.5	812	106	+0.2
+MS-209 (50 mg/kg)	6	10.0	10–11	111	+0.1
+MS-209 (100 mg/kg)	6	11.0	10–12	122	-0.2
+MS-209 (200 mg/kg)	6	13.0	1115	144	-0.1
Experiment 2					
control	12	9.0	911	100	+1.0
ADM (2 mg/kg)	6	11.0	10–12	122	-0.3
+MS-209 (200 mg/kg)	6	13.5	12-20	150	+0.3
+MS-209 (300 mg/kg)	6	17.5	13–21	194	-1.1
+MS-209 (450 mg/kg)	6	15.5	13–17	172	-1.9

^a CD2F₁ mice were given i.p. implants of 10⁶ P388/VCR leukemia cells on day 0. MS-209 was administered p.o. once a day just before i.p. injection of ADM from days 1 to 5.

^b Each value represents the mean of triplicate determinations. Standard deviations were within 10% of each value.

^c Numbers in parentheses represent relative values as compared with the IC₅₀ for each parent cell line in the absence of MS-209.

^b T/C value: median survival time of treated mice divided by median survival time of control mice.

^c Difference in body weight (g) between days 5 and 1.

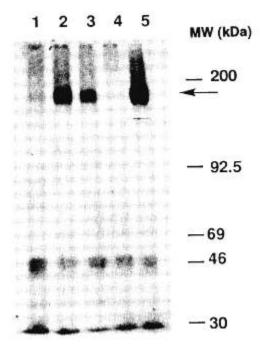


Figure 1. Inhibition of [3 H]azidopine photolabeling of P-glycoprotein. K562 (lane 1) and K562/ADM (lanes 2–5) membrane vesicles (50 μ g of protein) were incubated with 200 nM [3 H]azidopine in the absence (lanes 1 and 5) or presence of MS-209 at 1 (lane 2), 10 (lane 3) or 100 μ M (lane 4). After solubilization, photolabeled proteins were analyzed by sodium dodecyl sulfate–polyacrylamide gel electrophoresis.

suggest that MS-209 will be an orally-active MDR reversing drug.

mAbs

During the past 5 years, our group was developed immunotherapeutic approaches for overcoming MDR cancers using anti-P-glycoprotein mAbs, MRK16 and MRK17.47 We have evaluated the effect of MRK16 and MRK17 on the growth of human MDR ovarian cancer cells 2780AD. 48 2780AD cells (107/ mouse) were inoculated subcutaneously in athymic mice. In the experiment, we demonstrated growth inhibition of the mAbs. Animals received intravenous injection of various amounts of MRK16 or MRK17 on days 2 and 7 after tumor inoculation. Subcutaneous tumors never developed in the mice treated with 10 µg or more per animal of MRK16. When the mice were treated with 1 µg per animal of MRK16 or 1 or 10 µg per animal of MRK17, the development of tumors was substantially retarded as compared with control mice where palpable tumors developed by day 20.

In the next experiment, the therapy with antibodies was started after the tumors became palpable (150–400 mm³ in volume). One group of animals received 6 mg/kg of adriamycin intravenously three times as control therapy. Animals received various amounts of MRK16 or MRK17 intravenously on days 20, 23 and 26, and the average tumor volume was measured. At either 1 or 2 days after the first injection of MRK16, the size of the tumor masses remarkably decreased in all five mice. After the third MRK16 injection, the average tumor mass was substantially reduced and in four out of five mice the tumor mass had completely disappeared. Among these four mice, a palpable tumor recurred in one mouse on day 42; the other three mice remained tumor-free up to day 60. No significant inhibition of tumor growth was observed in animals which received 1 mg MRK17 intravenously on days 20, 23 and 26. These mAbs had no effect on the in vivo growth of drug-sensitive parent tumor A2780 cells, suggesting that the effect of these antibodies was specific for the drug-resistant tumor cells over-expressing the P-glycoprotein.

Several mechanisms, such as direct cytotoxicities of the mAbs or immune response induced by mAbs, may be involved in the growth inhibition of the xenografts. Complement-dependent cytotoxicity for 2780AD cells was clearly demonstrated *in vitro* using MRK16 and rabbit serum as complement. Antibody-dependent cell-mediated cytolysis by MRK16 was more clearly demonstrated than by MRK17. The benefit of MRK16 use *in vivo* in MDR cancer treatment has also been verified in the model of xenograft of MDR human colon cancer (HT29mdr1).⁴⁹

Conclusion

In this review, we have described therapeutic approaches to overcome MDR in cancer cells. Several new antitumor drugs were demonstrated to be effective for the treatment of resistant tumors *in vitro* and *in vivo*. As P-glycoprotein plays a central role in MDR, the therapeutic approaches targeting P-glycoprotein, such as application of chemosensitizing drugs and mAbs against P-glycoprotein, have been studied. Chemosensitizing drugs such as verapamil and cyclosporin A competitively inhibit the efflux of antitumor drugs from the cells by P-glycoprotein. Combination chemotherapy with antitumor agents and chemosensitizers could actually overcome clinical drug resistance in some refractory cancer patients. Several new chemosensitizing drugs have

been developed and some of them are under clinical phase study in Japan, the US and Europe. mAbs against P-glycoprotein are also attractive tools for overcoming MDR mediated by P-glycoprotein. Some antibodies, such as MRK-16, enhanced the immune responses to resistant tumors possessing P-glycoprotein. They also modulated the transport function of P-glycoprotein, and could be combined with antitumor drugs and chemosensitizers.

In the trials to reverse MDR mediated by P-glycoprotein, we must be concerned about the physiologic functions of P-glycoprotein expressed in various normal tissues. P-glycoprotein expressed in brain capillary endothelium could be functionally involved in the blood–brain barrier. P-glycoprotein in adrenal gland could be responsible for secretion of steroid hormones, P-glycoprotein in the luminal surface of the colon, brush border of proximal tubules in kidney and biliary canalicular surface of hepatocytes could contribute to the excretion of natural toxic substances into the lumen of the gastrointestinal tract, urine and bile, respectively. 55

Although the modulation of MDR is not easy to achieve, the therapeutic approaches described here are interesting. Further studies are needed to develop effective modalities to overcome drug resistance.

References

- 1. Tsuruo T. Mechanisms of multidrug resistance and implications for therapy. *Jpn J Cancer Res* 1987; **79**: 285–96.
- Roninson, IB, ed. Molecular and cellular biology of multidrug resistance in tumor cells. New York: Plenum 1989.
- 3. Safa AR, Glover CJ, Meyers MB, *et al.* Vinblastine photoaffinity labeling of a high molecular weight surface membrane glycoprotein specific for multidrug-resistant cells. *J Biol Chem* 1986; **261**: 6137–40.
- Cornwell MM, Safa AR, Felsted RL, et al. Membrane vesicles from multidrug resistant human cancer cells contain a specific 150- to 170-kDa protein detected by photoaffinity labeling. Proc Natl Acad Sci USA 1986; 83: 3847-50.
- Hamada H, Tsuruo T. Purification of the 170- to 180kilodalton membrane glycoprotein associated with multidrug resistance: the 170- to 180-kilodalton membrane glycoprotein is an ATPase. J Biol Chem 1988; 263: 1454– 8
- Hamada H, Tsuruo T. Characterization of the ATPase activity of the 170- to 180-kilodalton membrane glycoprotein (P-glycoprotein) associated with multidrug resistance. *Cancer Res* 1988; 48: 4926–32.
- Willingham MC, Richert ND, Cornwell MM, et al. Immunocytochemical localization of P170 at the plasma membrane of multidrug-resistant human cells. J Histochem Cytochem 1987; 35: 1451–6.

- 8. Shen D-W, Fojo A, Roninson IB, *et al.* Multidrug resistance in DNA-mediated transformants is linked to transfer of the human *mdr1* gene. *Mol Cell Biol* 1986; **6**: 4039–45.
- Gros P, Neriah YB, Croop JM, et al. Isolation and expression of a complementary DNA (mdr) that confers multidrug resistance. Nature 1986; 323: 728-31.
- Ueda K, Cardarelli C, Gottesman MM, et al. Expression of a full-length cDNA for the human mdr1 (P-glycoprotein) gene confer multidrug resistance. Proc Natl Acad Sci USA 1987; 84: 3004–8.
- 11. Bell DR, Gerlach JH, Kartner N, et al. Detection of P-glycoprotein in ovarian cancer: a molecular marker associated with multidrug resistance. J Clin Oncol 1985; 3: 311–5.
- 12. Fojo AT, Ueda K, Slamon DJ, *et al.* Expression of a multidrug resistance gene in human tumors and tissues. *Proc Natl Acad Sci USA* 1987; **84**: 265–9.
- Tsuruo T, Sugimoto Y, Hamada H, et al. Detection of multidrug resistance markers, P-glycoprotein and mdr1 mRNA, in human leukemia cells. Jpn J Cancer Res 1987; 78: 1415-9.
- Ishida Y, Ohtsu T, Hamada H, et al. Multidrug resistance in cultured human leukemia and lymphoma cell lines detected by a monoclonal antibody, MRK16. Jpn J Cancer Res 1989; 80: 1006–13.
- 15. Mizoguchi T, Yamada K, Furukawa T, et al. Expression of the MDR1 gene in human gastric and colorectal carcinomas. J Natl Cancer Inst 1990; 82: 1679–83.
- Pirker R, Wallner J, Geissler K, et al. MDR1 gene expression and treatment outcome in acute leukemia. J Natl Cancer Inst 1991; 83: 708-12.
- Holzmayer TA, Hilsenbeck S, Von Hoff D, et al. Clinical correlates of MDR1 (P-glycoprotein) gene expression in ovarian and small-cell lung carcinomas. J Natl Cancer Inst 1992; 84: 1486–91.
- Thiebaut F, Tsuruo T, Hamada H, et al. Cellular localization of the multidrug-resistance gene product P-glyco-protein in normal human tissues Proc Natl Acad Sci USA 1987; 84: 7735–8.
- Fairchild G, Ivy SP, Rushmore T, et al. Carcinogen-induced mdr overexpression is associated with xenobiotic resistance in rat preneoplastic liver nodules and hepatocellular carcinomas. Proc Natl Acad Sci USA 1987; 84: 7701-5.
- Sugawara I, Kataoka I, Morishita Y, et al. Tissue distribution of P-glycoprotein encoded by a multidrug resistant gene as revealed by a monoclonal antibody MRK16. Cancer Res 1988; 48: 1926–9.
- Sugawara I, Nakahama M, Hamada H, et al. Apparent stronger expression in the human adrenal cortex than in the human adrenal medulla of M_r 170,000–180,000 Pglycoprotein. Cancer Res 1988; 48: 4611–4.
- 22. Arceci RJ, Croop JM, Horwitz SB, et al. The gene encoding multidrug resistance is induced and expressed at high levels during pregnancy in the secretory epithelium of the uterus. Proc Natl Acad Sci USA 1988; 85: 4350-4.
- 23. Thiebaut F, Tsuruo T, Hamada H, et al. Immunohistochemical localization in normal tissues of different epitopes in the multidrug transport protein P170: evidence for localization in brain capillaries and crossreactivity of one antibody with a muscle protein. J Histochem Cytochem 1989; 37: 159–64.
- 24. Sugawara I, Hamada H, Tsuruo T, et al. Specialized

- localization of P-glycoprotein recognized by MRK16 monoclonal antibody in endothelial cells of the brain and the spinal cord. *Ipn J Cancer Res* 1990; **81**: 727–30.
- 25. Tsuruo T, Iida H, Tsukagoshi S, et al. Overcoming of vincristine resistance in P388 leukemia, in vivo and in vitro through enhanced cytotoxicity of vincristine and vinblastine by verapamil. Cancer Res 1981; 41: 1967–72.
- Rogan AM, Hamilton TC, Young RC, et al. Reversal of Adriamycin resistance by verapamil in human ovarian cancer. Science 1984; 224: 994-6.
- Tsuruo T, Iida H, Tsukagoshi S, et al. Increased accumulation of vincristine and Adriamycin in drug-resistant P388 tumor cells following incubation with calcium antagonists and calmodulin inhibitors. Cancer Res 1982; 42: 4730–3.
- 28. Tsuruo T, Iida H, Nojiri M, et al. Circumvention of vincristine and Adriamycin resistance in vitro and in vivo by calcium influx blockers. Cancer Res 1983; 43: 2905–10.
- Slater LM, Murray SL, Wetzel MM, et al. Verapamil restoration of daunorubicin responsiveness in daunorubicin-resistant Ehrlich ascites carcinoma. J Clin Invest 1982;
 10: 1131–4.
- Ramu A, Fuks Z, Gatt S, Glaubiger D. Reversal of acquired resistance to doxorubicin in P388 murine leukemia cells by perhexiline maleate. Cancer Res 1984; 44: 144–8
- 31. Ganapathi R, Grabowski D. Enhancement of sensitivity to Adriamycin in resistant P388 leukemia by the calmodulin inhibitor trifluoperazine. *Cancer Res* 1983; 43: 3696–9.
- 32. Tsuruo T. Circumvention of drug resistance with calcium channel blockers and monoclonal antibodies. In: Ozols RF, ed. *Drug resistance in cancer therapy.* New York: Kluwer 1989: 73–95.
- 33. Tsuruo T. Reversal of multidrug resistance by calcium channel blockers and other agents. In: Roninson IB, ed. *Molecular and cellular biology of multidrug resistance in tumor cells.* New York: Plenum 1991; 349–72.
- Naito M, Hamada H, Tsuruo T. ATP/Mg²⁺-dependent binding of vincristine to the plasma membrane of multidrug-resistant K562 cells. *J Biol Chem* 1988; 263: 11887– 91.
- Horio M, Gottesman MM, Pastan I. ATP-dependent transport of vinblastine in vesicles from human multidrugresistant cells. Proc Natl Acad Sci USA 1988; 85: 3580-4.
- Safa AR. Photoaffinity labeling of the multidrug-resistance-related P-glycoprotein with photoactive analogs of verapamil. *Proc Natl Acad Sci USA* 1988; 85: 7187–91.
- 37. Yusa K, Tsuruo T. Reversal mechanisms of multidrug resistance by verapamil: direct binding of verapamil to P-glycoprotein on specific sites and transport of verapamil outward across the plasma membrane of K562/ADM cells. *Cancer Res* 1989; **49**: 5002–6.
- Slater LM, Sweet P, Stupeckey M, et al. Cyclosporin A reverses vincristine and daunorubicin resistance in acute lymphatic leukemia in vitro. J Clin Invest 1986; 77: 1405– 8.
- Natio M, Tsuruo T. Competitive inhibition by verapamil of ATP-dependent high affinity vincristine binding to the plasma membrane of multidrug-resistant K562 cells without calcium ion involvement. *Cancer Res* 1989; 49: 1452– 5.
- 40. Sonneveld P, Durie BGM, Lokhorst HM, et al. Modulation

- of multidrug-resistant multiple myeloma by cyclosporin. *Lancet* 1992; **340**: 255–9.
- Sato W, Fukazawa N, Suzuki T, et al. Circumvention of multidrug resistance by a newly synthesized quinoline derivative, MS-073. Cancer Res 1991; 51: 2420-4.
- Naito M, Oh-hara T, Yamazaki A, et al. Reversal of multidrug resistance by an immunosuppressive agent FK-506. Cancer Chemother Pharmacol 1992; 29: 195-200.
- Boesch D, Muller K, Manzanedo AP, et al. Restoration of daunomycin retention in multidrug-resistant P388 cells by submicromolar concentrations of SDZ PSC 833, a nonimmunosuppressive cyclosporin derivative. Exp Cell Res 1991; 196: 26–32.
- Boesch D, Gaveriaux C, Jachez B, et al. In vivo circumvention of P-glycoprotein-mediated multidrug resistance of tumor cells with SDZ PSC 833. Cancer Res 1991; 51: 4226–33.
- 45. Watanabe T, Tsuge H, Oh-hara T, et al. Comparative study on reversal efficacy of SDZ PSC 833, cyclosporin A and verapamil on multidrug resistance in vitro and in vivo. Acta Oncol 1994; in press.
- Sato W, Fukazawa N, Nakanishi O, et al. Reversal of multidrug resistance by a novel quinoline derivative, MS-209. Cancer Chemother Pharmacol 1994; in press.
- Hamada H, Tsuruo T. Functional role for the 170- to 180kDa glycoprotein specific to drug-resistant tumor cells as revealed by monoclonal antibodies. *Proc Natl Acad Sci* USA 1986; 83: 7785–9.
- Tsuruo T, Hamada H, Sato S, Heike Y. Inhibition of multidrug-resistant human tumor growth in athymic mice by anti-P-glycoprotein monoclonal antibodies. *Jpn J Cancer Res* 1989; 80: 627–31.
- 49. Pearson JW, Fogler WE, Volker K, et al. Reversal of drug resistance in a human colon cancer xenograft expressing MDR1 complementary DNA by a in vivo administration of MRK-16 monoclonal antibody. J Natl Cancer Inst 1991; 83: 1386–91.
- Tatsuta T, Naito M, Oh-hara T, et al. Functional involvement of P-glycoprotein in blood-brain barrier. J Biol Chem 1992; 267: 20383–91.
- Tsuji A, Terasaki T, Takabatake Y, et al. P-glycoprotein as the drug efflux pump in primary cultured bovine brain capillary endothelial cells. Life Sciences 1992; 51: 1427– 37.
- Shirai A, Naito M, Tatsuta T, et al. Transport of cyclosporin A across the brain capillary endothelial cell monolayer by P-glycoprotein. Biochim Biophys Acta 1994; 1222: 400–4.
- Naito M, Yusa K. Tsuruo T. Steroid hormones inhibit binding of *vinca* alkaloid to multidrug resistance related P-glycoprotein. *Biochem Biophys Res Commun* 1989; 158: 1066-71.
- 54. Yang C-PH, DePinho SG, Greenberger LM, et al. Progesterone interacts with P-glycoprotein in multidrug-resistant cells and in the endometrium of gravid uterus. J Biol Chem 1989; 264: 782–8.
- Kamimito Y, Gatmaitan Z, Hsu J, et al. The function of Gp170, the multidrug resistance gene product, in rat liver canalicular membrane vesicles. J Biol Chem 1989; 264: 11693–8.

(Received 3 November 1994; accepted 15 December 1994)